

The water quality parameters controlling diatoms assemblage in Rawapening Lake, Indonesia

T.R. SOEPROBOWATI^{1,2,*}, S.D. TANDJUNG³, SUTIKNO⁴, S. HADISUSANTO³, P. GELL⁵, HADIYANTO^{6,**}, S.W.A. SUEDY²

¹School of Postgraduate Studies, Universitas Diponegoro. Semarang 50241, Central Java, Indonesia

²Department of Biology, Faculty of Science and Mathematics, Universitas Diponegoro. Jl. Prof. Soedarto, SH, Kampus Tembalang, Semarang 50275, Central Java, Indonesia. Tel./Fax.: +62-24-70799494, *email: trsoeprbowati@live.undip.ac.id

³Faculty of Biology, Universitas Gadjah Mada, Sleman 55281, Yogyakarta, Indonesia

⁴Faculty of Geography, Universitas Gadjah Mada, Sleman 55281, Yogyakarta, Indonesia

⁵School of Applied and Biomedical Sciences, Faculty of Science and Technology, Federation University, Australia

⁶Department of Chemical Engineering, Universitas Diponegoro. Semarang 50275, Central Java, Indonesia. **email: hadiyanto@live.undip.ac.id

Manuscript received: 16 May 2016. Revision accepted: 11 August 2016.

Abstract. Soeprbowati TR, Tandjung SD, Sutikno, Hadisusanto S, Gell P, Hadiyanto. 2016. The water quality parameters controlling diatoms assemblage in Rawapening Lake, Indonesia. *Biodiversitas* 17: 657-664. Diatoms are microalgae that have an important role in aquatic ecosystem. The silicious diatoms fossil had been used for paleolimnological analysis. However, data set diatoms of Indonesia does not develop yet. The aims of this research were to assess water quality parameters controlling diatoms assemblage of Rawapening Lake, to assess transfer function, to develop diatoms data set of Rawapening, and to compare Rawapening diatoms data set with European diatoms data set for the reconstruction of total phosphorous and pH. Water samples were taken from 3 sites of Rawapening Lake for water quality analysis. Sediment samples had been taken from 3 sites in Rawapening Lake using hand auger, and were sliced every 0.5 cm for diatoms analysis. Diatoms slides were prepared from about 5 g of dry sediment using 10% of chloride acid followed by 10% of peroxide to remove organic matters and carbonates, respectively. 300 diatom valves were calculated on the entire samples, calculation the number of valve's diatom aims to identify the lowest level of taxonomy. Phosphate, temperature, and calcium were the environmental parameters that influence the diatoms assemblage of Rawapening Lake. Phosphate was contributed 50% on diatoms assemblage. Internal based transfer function of diatoms provides more suitable diatoms data set rather than European diatom data set. The Rawapening diatoms data set of total phosphorous is the initial Indonesian diatoms data set for past trophic status, therefore, research has to be continued spatially for other Indonesian lakes and temporary on specific Indonesian lake to develop Indonesia diatoms data set.

Keywords: Diatoms, Indonesia, phosphate, Rawapening Lake, water quality

INTRODUCTION

Global concern on the environmental issue had been focused on the sustainability of the availability, management and conservation of freshwater ecosystem. The phenomenon of increasing population growth, urbanization, wastewater management, will influence water quality problem in the future, particularly in Asia, therefore required an effort to overcome the problem of water quality degradation (Avans et al. 2012).

Freshwater quality problem were also occurred in Indonesia, and become a national problem. In 2009 there was an agreement between 9 Indonesian ministers about 7 lakes priority criteria, regarding to the lake degradations: (i) sedimentation; (ii) pollution, eutrophication, and water quality degradation; (iii) lake usage: power electricity, agriculture, fisheries, drinking water, social and religious life, tourism; (iv) commitment between government and community; (v) lake strategy for national function; (vi) biodiversity (endemic species); and (vii) level of risk disaster. There were 7 programs for 15 lakes priority, set about lake ecosystem management, scientific and technological approach on using lakes resources; the

development of lake monitoring, evaluation and information systems; preparation of adaptation and mitigation steps; the impact of environmental changes to the lakes; the development of capacity, regulation and coordination; improvement of community involvement; and sustainable funded. There are 15 lakes national priority in the year 2010-2014 that are Lakes of Toba Maninjau, Singkarak, Kerinci, Tondano, Limboto, Poso, Tempe, Matano, Cascade Mahakam Semayang-Melintang-Jempang, Sentarum, Sentani, Rawa Danau, Batur, and Rawapening (MoE 2010).

Rawapening Lake is located in the S 7°04'-7°30' and E 110°24'46"-110°49'06" in the urban area between of 3 cities in the Central Java, i.e.: Yogyakarta, Solo, and Semarang. Despite of its small size (26.23 km²), 207,438 people depend their life to Rawapening Lake. The catchment area of the lake is consists of 55.40 km² paddy's fields; 1,126.42 km² of plantations; 44.08 km² of settlements; and 21.64 km² of forests (BPS 2010). Ecologically, Rawapening Lake had been changed particularly because there are plants that grow uncontrollably due the influence of eutrophication, 70% of lake area are covered by water hyacinth. Eutrophication

and sedimentation were huge problem that had induced lake shallowest. This environmental degradation had threatened its function for electricity power, irrigation, fisheries, source of drinking water and recreation area. These were typical problem of Indonesian lakes. Limnological research was 1 out of 6 super priority programs for Save Rawapening Lake have to be developed as a basic national policy (MoE 2011).

It was not doubtly about the potential use of diatoms for bioindicator of water quality, and used diatoms to reconstruct past water quality changes (Gell et al. 2007; Reid and Ogden 2009; Soeprbowati et al. 2012; Adams et al. 2014; Yun et al. 2014). Diatoms are dominant microalgae almost in all aquatic ecosystems, contribute 20-25% primary production, have an important role in the silica and carbon cycle (Soeprbowati et al. 2012). Different taxa have different toleration to the environmental parameters. Therefore, diatoms assemblage reflects water quality effectively. Quantitatively, reconstruction of past condition can be done by 3 approaches namely indicator species approach, involving bioclimatic modeling that was causal relationship between species distribution and climate variable; assemblage approach involving modern analog technique and response surfaces; and multivariate calibration-functional that known as a transfer function have an important role in future quaternary paleoecology (Birks et al. 2010).

The aims of this research were to assess water quality parameters controlling diatoms assemblage of Rawapening Lake Indonesia, to assess transfer function and to develop diatoms data set of Rawapening, and to compare Rawapening diatoms data set with European diatoms data set on reconstruction of total phosphorous and pH.

MATERIALS AND METHODS

Three research sites were chosen based on representative of inlet, outlet, and water body of the Rawapening Lake, Central Java, Indonesia (Figure 1). Hand-auger was used to collect sediment samples, and then slice every 0.5 cm thick for diatoms analysis. 5 gr of dry sediment was digested by heated with 10% of chloride acid followed by 10% of peroxide for 2 hours to remove organic matters and carbonates, respectively (Battarbee et al. 2001). Distilled water was added when the solution almost dry. After settled for at least 4 hours, supernatant was discharged and added with distilled water until 50 mL and allowed to settle for another 4 hours. This was done repeatedly until the pH was neutral (7). The washed frustules were mounted in Naphrax and examined by optical microscope at 1,000 magnifications. In each sample, an average of 300 valves was counted in order to establish the relative abundance of the species (Soeprbowati 2010). Diatom taxonomy followed the guidelines Kramer and Lange-Bertalot (2010a,b (teil 1-2), 2008 (teil 3), 2011 (teil 4) volume 1-4; Guiry and Guiry 2016) and valves were identified to the lowest taxonomic level possible.

The water quality measurements was done in-situ by measuring the temperature, pH, dissolved oxygen, turbidity, conductivity, and water clarity on the depth of 20 cm from surface water. The Rawapening lake depth was measure from the surface water until the bottom of the lake. Water samples were also taken to analyze the concentration of dissolved silicate, total suspended solid (particulate material), biological oxygen demand (BOD), calcium, Fe, Mg, Na, Pb, Cd, Cu, Cr, phosphate, total phosphorous, total nitrogen, nitrite, nitrate, ammonia, and chlorophyll-*a*. BOD was measure by counted of dissolved oxygen on the first day and 5 days. Total phosphorous and total nitrogen was the total of minerals and organics part. 40 data from 2004-2008 had been compiled to develop training set of Rawapening (RP40), that were chosen for the same parameter from the same site as time series. Water quality variables were transformed by $\log(x+1)$.

Principle Component Analysis (PCA) followed by Canonical Component Analysis (CCA) was done to determine the water quality parameters that influence the diatoms assemblage and its distribution. CANOCO version 4.56 (ter Braak and Smilauer 2009) was used to do PCA and CCA. CCA was designed to detect the variation trend of diatoms species due to environmental variables, the CCA had combine regular ordination aspect and regression (Kireta et al. 2012). The relationship between diatoms distribution and environmental dataset was assessed by direct gradient technique by CCA with the limited axis ordination as a linier combination of environmental variables (ter Braak and Smilauer 2009). Forward selection was used to reduce redundancy variables and select significantly variables that able to reflect the variation on the diatoms data set by unlimited Monte Carlo permutation. The variables that significantly responsible to the diatoms distribution are when $p < 0.05$ on the 199 permutation. Insignificant variables were not included in the further analysis. Samples of data set as well as environmental parameters were selected. The samples with inflation variant > 20 have a stable data effect, so those samples did not include in the analysis (ter Braak and Smilauer 2009).

Furthermore, transfer function was analyzed with Computer Program C2 1.7.6 (Juggins 2014). The diatoms species with more than 5% of relative abundance had been included in the analysis with 3 significant environmental parameters, i.e. phosphate, calcium, and temperature. *Weighted Averaging* (WA) with inversed and deshrinking was the model that implemented for multivariate calibration function to estimate water quality based on diatoms (Birks et al. 2010). Prediction model was assessed based on correlation coefficient (R^2) between observed diatoms and inference diatoms in the form of Root Mean Square Error (RMSE). The validation for prediction error had assessed with leave-one-out jackknifing (Juggins 2014).

Reconstruction past condition of nutrient status of the lake was done based on the European diatom data set with Computer Program of ERNIE (Environmental Reconstruction using the European Diatom Database (Juggins 2016), compare to Rawapening diatom data set.

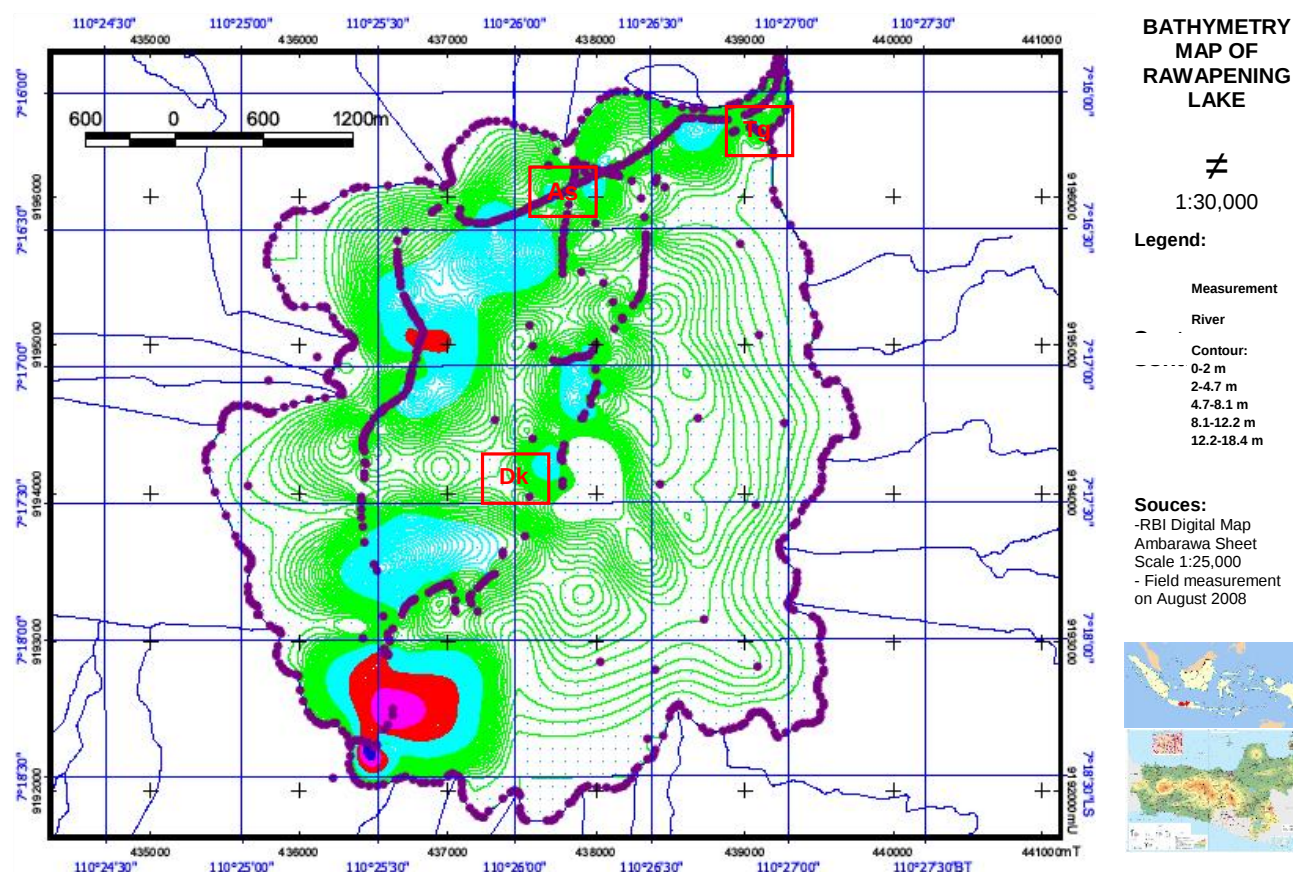


Figure 1. Research site for water quality and diatoms in Rawapening Lake, Central Java, Indonesia

Table 2. Marginal and conditional effect of environmental variables on the diatom variation in Rawapening Lake, Indonesia

Variable	Marginal effects		Conditional effects			
	Var. N	Lambda1	Var. N	LambdaA	P	F
Phosphate	13	0.36	13	0.36	0.005	3.51
Calcium	23	0.32	23	0.3	0.005	3.03
Temperature	3	0.3	3	0.16	0.001	1.7
Turbidity	27	0.35	27	0.13	0.06	1.37
Silica	10	0.3	10	0.12	0.125	1.28
Conductivity	2	0.16	2	0.12	0.175	1.21
Fe	26	0.24	26	0.1	0.365	1.12
Water clarity	4	0.14	4	0.1	0.27	1.1
Depth	5	0.16	5	0.11	0.225	1.2
pH	1	0.32	1	0.1	0.5	1.01
Mg	24	0.3	24	0.09	0.445	1
TSS	17	0.15	17	0.09	0.55	0.92
Pb	6	0.08	6	0.08	0.56	0.9
Nitrite	16	0.18	16	0.08	0.605	0.88
Nitrate	15	0.22	15	0.09	0.56	0.95
Total phosphorous	11	0.14	11	0.08	0.68	0.79
Cd	7	0.09	7	0.07	0.695	0.73
Cr	8	0.08	8	0.12	0.275	1.29
Cu	9	0.08	9	0.12	0.235	1.27
BOD	29	0.19	29	0.06	0.76	0.7
Na	25	0.28	25	0.09	0.57	0.9
Chlorophyll	20	0.27	18	0.08	0.64	0.83
NH ₃	14	0.31	14	0.06	0.74	0.67
Dissolved oxygen	28	0.23	28	0.06	0.93	0.55
Total nitrogen	12	0.25	12	0.11	0.405	1.09

RESULTS AND DISCUSSION

The Rawapening diatoms data set was started by Principle Component Analysis (PCA) on the water quality, followed by Canonical Correspondence Analysis (CCA). PCA is a multivariate analysis with an ordination technique which developed theoretical variables by minimized total residual after fitted the strike line and regression (Jongman et al. 2005). Using 21 environmental parameters from 40 sites compiling with previous research (2004-2008) in Rawapening Lake, the Eigen value to axis 1 and 2 were 0.417 and 0.108, which reflected 65.2% environmental variation (Table 1). Axis 1 was dominated by phosphate and temperature, whereas axis 2 was connected with more heterogeneous variables. The environmental variable was significantly influence the diatoms assemblage if $p < 0.05$ with the permutation 199. Temperature and phosphate were in the first axis, whereas calcium was in the fourth axis (Figure 2). Phosphate explained 41.7% species variation on the Rawapening diatoms data set whereas calcium explained 65.2% species variation in Rawapening (Table 1).

All environmental variables, consisting of temperature, pH, dissolved oxygen, turbidity, conductivity, water clarity, dissolved of silicate, total suspended solid (particulate material), BOD, calcium, Fe, Mg, Na, Pb, Cd, Cu, Cr, phosphate, total phosphorous, total nitrogen, nitrite, nitrate,

ammonia, and diatoms data set from 40 sites were analyzed. Furthermore, based on the Monte Carlo permutation test, the environmental variables from marginal effect with $\lambda > 0.2$ were reduced from the data. However, based on conditional effect, 3 variables that responsible to the diatoms assemblage ($p < 0.05$) were phosphate, calcium, and temperature (Table 2). Those result related to the allochthonous effect from the inlet.

Based on the CCA, the variation of Rawapening's diatoms was controlled by phosphate, temperature and calcium (Figure 2). The diatom species that correlate to the environmental parameters were provided in Table 3. Phosphate was the strongest variable that influenced diatoms assemblage in Rawapening Lake with λ of 0.36 and $p < 0.005$ (Table 2).

Calcium carbonate is the most contributors for water alkalinity. Alkalinity was buffer capacity for water pH. Bicarbonate was alkali due to the reaction with H^+ , meanwhile when hydrolyzed it may produce OH^- . The alkalinity of CO_3^{2-} were stronger than CO_2 , therefore in equilibrium condition, OH^- in the bicarbonate solution always exceed H^+ . Based on this research, the major cation dominated in Rawapening was calcium. People living surrounding Rawapening were used calcium carbonate on the production of organic fertilizer from water hyacinth. The tailing from fertilizer process diffuses to Rawapening Lake. Diatoms used bicarbonate ion as source of carbon and induced hydroxide accumulation ($CO_3^{2-} + H_2O \leftrightarrow HCO_3^- + OH^-$). In turn, this hydroxide accumulation may induce an increase of pH (9-10). This alkali condition had also indicated by blooming of *Aulacoseira granulata* (Ehr.) Simonsen (Soeprbowati et al. 2012).

Development of transfer function basically was based on the Weighted Averaging (WA) with inversed and deshrinking. WA calibration was more suitable rather than WAT (Weighted Averaging with Tolerance-down weighting) as a predictive model of phosphate. Diatoms assemblage was correlated with phosphate ($R^2 = 0.50$, RMSE = 0.22, Table 4). Phosphate contributes 50% on diatom assemblage, and the rest were another factors.

Generally, diatom data set and transfer function was developed based on some lakes numbers. Mills (2009) developed transfer function based on 86 Uganda's lakes. However, Mackay et al. (2003) had successfully developed transfer function based on the internal diatoms of Baikal Lake. The exploration of quantitative correlation between diatom species and environmental variables had been calculated based on the environmental internal gradient of Rawapening Lake and shown a quite good trend.

Table 1. Eigen value 4 axis resulting from PCA

Axis	1	2	3	4	Total variant
Eigen value	0.417	0.108	0.067	0.059	1.0
Correlation species-environment	0.853	0.829	0.793	0.826	
% cumulative species data	41.7	52.6	59.3	65.2	
Sum of Eigen value = 1					
Sum of canonical Eigen value = 0.681					

The reconstruction of total phosphorous of Rawapening Lake based on the European diatoms data set compare to Rawapening diatoms data set shown a similar pattern for Asinan and Tuntang sites (Figure 3). There was an increase of total phosphorous in between 1984-1990 which was related to the low of rainfall. Total phosphorous from catchment area were entered to Rawapening Lake from 16 inlets, whereas the only one outlet did not discharge the water during the dry season to maintain water level for electricity power. This condition had induced a sharp increase of total phosphorus content.

However, the reconstruction from Dangkel site shown a different pattern. There was a total phosphorous concentration decreasing trend in the middle layers with European diatoms data set, but an increase trend with Rawapening diatoms data set (Figure 3). For Dangkel site, total phosphorous content more representatively reconstruct with Rawapening diatoms data set rather than European data set, due to diatoms variation species at Dangkel site which was higher than European data set, which are not represented in European diatoms data set.

The reconstruction of pH of Rawapening lake had differ patten for different sites both using European diatoms data set and Rawapening diatoms data set. For examples, Asinan site shown fluctuate pH when reconstruct by European diatoms data set. However, based on Rawapening diatoms data set, the Rawapening pH tends to increase by year (Figure 4). Many diatoms species found in Rawapening Lake, do not found in European diatoms data set. In Rawapening diatoms data set, those species characterized such specific condition.

The pH of Rawapening Lake was fluctuant. In 1978, pH in Rawapening was neutral about 7.2-7.6 (Goltenboth 1979). In 1999, pH tent to increase, in the range of 7.5-8.8 (BPDL-PPLH Undip 1999). In 2003, pH relatively neutral, in the range of 6.5-7.7 (Wibowo 2004). In 2004 and 2005, pH in the inlet rivers tent to neutral, but in the site near spring and floating island, the pH tent to be high which was 9.52 (Soeprbowati et al. 2012). In this research (2009), the pH of Rawapening Lake was neutral, except near the spring and floating island that were 11 and 9.2. Naturally, lake will be more acid by time, but in Rawapening as stated before, the pH tent to increase. This condition is caused by the decomposition of organic materials by microbes in the peat sediment. The other reason is the use of calcium for treat organic fertilizer from water hyacinth. In the rainy day, the calcium leaches and enters to the lake. In recent study, those paleoreconstruction of ecological change in Rawapening was proven a trend of increasing pH. In the measurement in June 2015, pH of Asinan site was 7.8-8.1 (Soeprbowati 2015).

Internal based diatoms of Rawapening Lake developed from different 40 sites and time scale, shown quite good results since it was usually represent the specific diatoms. The diatoms data set of Rawapening was initial Indonesian diatoms data set for reconstruction total phosphorous, therefore, research has to be continued to add and fulfill data of water quality and diatoms from other Indonesian lakes.

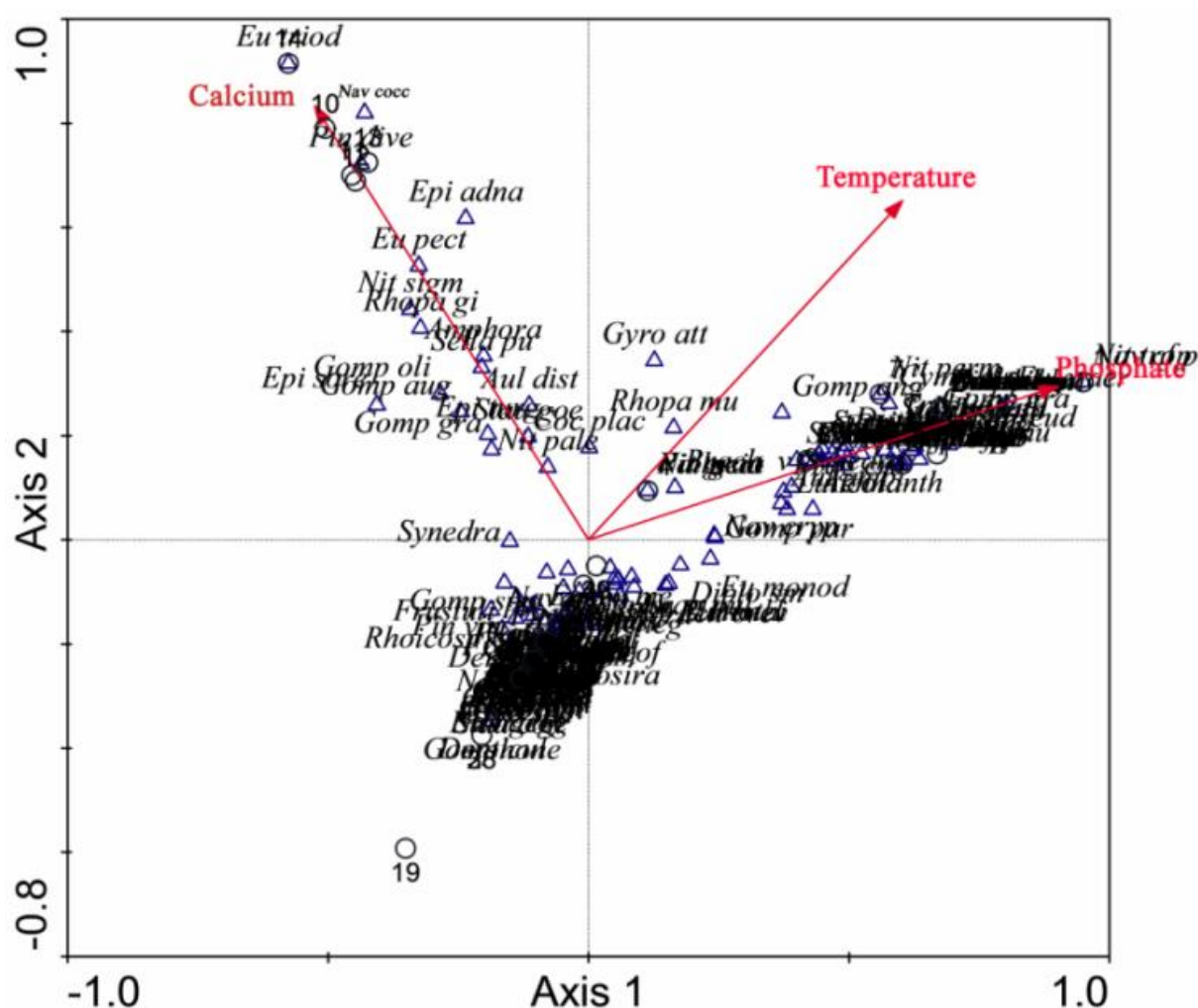


Figure 2. Triplot CCA; Phosphate, temperature and calcium the major of environmental variables which influence diatom variation of Rawapening Lake, Central Java, Indonesia

Table 3. List of diatom species of Rawapening Lake, Central Java, Indonesia in correlation with water quality as mention in Figure 2.

Abbreviation	Name of species		
Ach brev	<i>Achnanthes brevipes</i> Agardh	Camp nor	<i>Campylodiscus noricus</i> Ehrenberg ex Kutzing
Ach exi	<i>Achnantheidium exiguum</i> (Grunow)	Coc pla	<i>Cocconeis placentula</i> Ehrenberg
Ach exi1	<i>Achnantheidium exiguum1</i> Grunow	Cy atom	<i>Cyclotella atomus</i> Hustedt
Ach inf	<i>Achnantheidium inflata</i> (Kutzing) Grunow	Cy men	<i>Cyclotella meneghiniana</i> Kutzing
Ach min	<i>Achnantheidium minutissimum</i> (Kutzing) Czamecki	Cy pse	<i>Cyclotella pseudostelligera</i> Hustedt
Ach neo	<i>Achnantheidium neomicrocephalum</i> Kutzing	Cym pros	<i>Cymbella prostata</i> (Berkeley) Cleve
Amp cop	<i>Amphora copulata</i> (Kutzing) Schoeman & R.E.M. Archibald	Cym sol	<i>Cymatopleura solea</i> (Brebisson) W Smith
Amp hol	<i>Amphora holsatica</i> W. Smith	Cymb af	<i>Cymbella affinis</i> Kutzing
Amp ova	<i>Amphora ovalis</i> (Kutzing) Kutzing	Cymb cae	<i>Cymbella caespitosum</i> Kutzing
Amp sp	<i>Amphora</i> sp.	Cymb cis	<i>Cymbella cistula</i> (Ehrenberg) O. Kirchner
Amp ven	<i>Amphora veneta</i> Kutzing	Cymb tum	<i>Cymbella tumida</i> (Brebisson) Van Heurck
Amph ala	<i>Amphipora alata</i> (Ehrenberg) Kutzing	Den ele	<i>Denticula elegans</i> Kutzing
Amph cof	<i>Amphora coffeaeformis</i> Kutzing	Den sp	<i>Denticula</i> sp.
Ano sphae	<i>Anomoeoneis sphaerophora</i> Pfitzer	Dia cont	<i>Diademesmis contenta</i> (Grunow) D.G. Mann
Aul ambi	<i>Aulacoseira ambigua</i> (Grunow) Simonsen Manitoba	Dia conf	<i>Diademesmis confervacea</i> Kutzing
Aul dis	<i>Aulacoseira distans</i> (Ehrenberg) Simonsen	Dia hie	<i>Diatoma hiemale</i> (Roth) Heiberg
Aul gran	<i>Aulacoseira granulata</i> (Ehrenberg) Simonsen	Dia vul	<i>Diatoma vulgare f. brevis</i> (Grunow) Bukhtiyarova
Bac pax	<i>Bacillaria paxillifer</i> (O. F. Muller) T. Marsson	Diplo el	<i>Diploneis elliptica</i> (Kutzing) Cleve
Brach breb	<i>Brachysira brebissonii</i> Ross	Diplo ova	<i>Diploneis ovalis</i> (Hilse) Cleve
Brach neo	<i>Brachysira neoxilis</i> Lange-Bertalot	Diplo smi	<i>Diploneis smithii</i> (Brebisson) Cleve
Brach vit	<i>Brachysira vitrea</i> Grunow	Disc stel	<i>Discostella stelligera</i> (Cleve & Grunow) Houk & Klee
Calo ven	<i>Caloneis ventricosa</i> (Eggenberg) F. Meister	Ency mes	<i>Encyonema mesiana</i> (Cholnoky) DGMann
		Eny min	<i>Encyonema minutum</i> (Hilse) DGMann
		Epi ad	<i>Epithemia adnata</i> (Kutzing) Brebisson
		Epi ar	<i>Epithemia argus</i> Kutzing
		Epi tur	<i>Epithemia turgida</i> (Ehrenberg) Kutzing

Eu bi	<i>Eunotia bilunaris</i> (Ehrenberg) Schaarschmidt	Nav rad	<i>Navicula radiosa</i> Kutzing
Eu cam	<i>Eunotia camelus</i> Ehrenberg	Nav rhyn	<i>Navicula rhyncocephala</i> Kutzing
Eu exi	<i>Eunotia exigua</i> (Brebisson ex Kutzing) Rabenhorst	Nav spp	<i>Navicula</i> spp.
Eu inc	<i>Eunotia incisa</i> W. Smith ex W Gregory	Nei aff	<i>Neidium affine</i> (Ehrenberg) Pfitzer
Eu mon	<i>Eunotia monodon</i> Ehrenberg	Nit aci	<i>Nitzschia acicularis</i> (Kutzing) W. Smith
Eu nae	<i>Eunotia naegelli</i> Migula	Nit cap	<i>Nitzschia capitellata</i> Hustedt
Eu par	<i>Eunotia parallela</i> Ehrenberg	Nit clau	<i>Nitzschia clausii</i> Hantzsch
Eu pec var pec	<i>Eunotia pectinalis</i> var. <i>pectinalis</i> Kutzing	Nit clos	<i>Nitzschia closterium</i> (Ehrenberg) W. Smith
Eu prae	<i>Eunotia praeupta</i> Ehrenberg	Nit dis	<i>Nitzschia dissipata</i> (Kutzing) Rabenhorst
Eu sep	<i>Eunotia septentrionalis</i> Geniculata .Berg	Nit fil	<i>Nitzschia filiformis</i> (W. Smith) Van Heurck
Eu ten	<i>Eunotia tenella</i> (Grunow) Hustedt	Nit gra	<i>Nitzschia gracilis</i> Hantzsch
Eu trio	<i>Eunotia triodon</i> Ehrenberg	Nit lin	<i>Nitzschia linearis</i> W. Smith
Fal pyg	<i>Fallacia pygmaea</i> (Kutzing) Stickle & D.G.Mann	Nit pal	<i>Nitzschia palea</i> (Kutzing) W. Smith
Frag cap	<i>Fragilaria capitata</i> (Ehrenberg) Lange-Bertalot	Nit per	<i>Nitzschia perminuta</i> (Grunow) M. Peragallo
Frag capu	<i>Fragilaria capucina</i> Desmazieres	Nit sig	<i>Nitzschia sigmoidea</i> (Nitzsch) W. Smith
Frag capu var rum	<i>Fragilaria capucina</i> var. <i>rumpens</i> (Kutzing) Lange-Bertalot	Nit trop	<i>Nitzschia tropica</i> Hustedt
Frag croto	<i>Fragilaria crotonensis</i> Kitton	Ortho ros	<i>Orthoseira roseana</i> (Rabbenh.) O'Meara
Frag int	<i>Fragilaria intermedia</i> Grunow	Pin bor	<i>Pinnularia borealis</i> Ehrenberg
Frag pin	<i>Fragilaria pinnata</i> Ehrenberg	Pin brau	<i>Pinnularia braunii</i> Cleve
Frag pul	<i>Fragilaria pulchella</i> (Ralfs ex Kutzing) Lange-Bertalot	Pin div	<i>Pinnularia divergentissima</i> (Grunow) Cleve
Frag vau	<i>Fragilaria vaucheriae</i> (Kutzing) J. B. Petersen	Pin gen	<i>Pinnularia gentilis</i> (Donkin) Cleve
Frag vir	<i>Fragilaria virescens</i> Ralfs	Pin gib	<i>Pinnularia gibba</i> Ehrenberg
Frus rhom	<i>Frustulia rhomboides</i> (Ehrenberg) De Toni	Pin int	<i>Pinnularia interrupta</i> W. Smith
Frus vul	<i>Frustulia vulgaris</i> (Thwaites) De Toni	Pin meso	<i>Pinnularia mesolepta</i> (Ehrenberg) W. Smith
Gomp acu	<i>Gomphonema acuminatum</i> Ehrenberg	Pin micro	<i>Pinnularia microstauron</i> (Ehrenberg) Cleve
Gomp af	<i>Gomphonema affine</i> Kutzing	Pin neo	<i>Pinnularia neo-major</i> Krammer
Gomp ang	<i>Gomphonema angustum</i> C. Agardh	Pin nob	<i>Pinnularia nobilis</i> Ehrenberg (Ehrenberg)
Gomp aug	<i>Gomphonema augur</i> Ehrenberg	Pin sp	<i>Pinnularia</i> sp.
Gomp clav	<i>Gomphonema clavatum</i> Ehrenberg	Pin sto	<i>Pinnularia stomatophora</i> Grunow
Gomp grac	<i>Gomphonema gracilis</i> Hustedt	Pin subcap	<i>Pinnularia subcapitata</i> W. Gregory
Gomp grac tur	<i>Gomphonema gracilis turris</i> Hustedt	Pin subgib	<i>Pinnularia subgibba</i> Krammer
Gomp oli	<i>Gomphonema olivaceum</i> (Hornemann) Brebisson	Pin supdiv	<i>Pinnularia superdivergentissima</i> Chaumont & H. Germain
Gomp par	<i>Gomphonema parvulum</i> (Kutzing) Kutzing	Pin vir	<i>Pinnularia viridis</i> (Nitzsch) Ehrenberg
Gomp spp	<i>Gomphonema</i> spp.	Pla del	<i>Planothidium delicatulum</i> Kutzing
Gomp trun	<i>Gomphonema truncatum</i> Ehrenberg	Pla lan	<i>Planothidium lanceolatum</i> Brebisson
Gomp ven	<i>Gomphonema ventricosum</i> W. Gregory	Pleu	<i>Pleurosira</i> sp.
Gyro acu	<i>Gyrosigma acuminatum</i> Kutzing Rabenhorst	Psam didy	<i>Psammothidium didymium</i> Hustedt
Gyro at	<i>Gyrosigma attenuatum</i> (Kutzing) Rabenhorst	Rhoi abb	<i>Rhoicosphenia abbreviata</i> (C. Agardh) Lange-Bertalot
Gyro obt	<i>Gyrosigma obtusatum</i> (Sullivant & Wormley) C.S Boyer	Rhoi cur	<i>Rhoicosphenia curvata</i> (Kutzing) Grunow
Gyro par	<i>Gyrosigma parkerii</i> (M.B. Harrison) Boyer	Rhop gib	<i>Rhopalodia gibba</i> (Ehrenberg) Otto Muller
Gyro scal	<i>Gyrosigma scalproides</i> (Rabenhorst) Cleve	Rhop mus	<i>Rhopalodia musculus</i> (Kutzing) Otto Muller
Gyro spen	<i>Gyrosigma spencerii</i> (Bailey ex Quekett) Griffth & Henfrey	Sel pup	<i>Sellaphora pupula</i> (Kutzing) Mereschkovsky
Han arc	<i>Hannaea arcus</i> (Ehrenberg) R.M. Patrick	Sel sem	<i>Sellaphora seminulum</i> (Grunow) D.G. Mann
Hantz amp	<i>Hantzschia amphioxys</i> (Ehrenberg) Grunow	Sel wis	<i>Sellaphora wislouchii</i> Poretzky
Lut hal	<i>Luticola halophila</i> Grunow	Stau an var jav	<i>Stauroneis anceps</i> var. <i>javanica</i> Hustedt
Lut mut	<i>Luticola mutica</i> (Kutzing) D.G. Mann	Stau coh	<i>Stauroneis cohnii</i> Hilse
Mas el	<i>Mastogloia elliptica</i> (C. Agardh) Cleve	Stau cons	<i>Stauroneis construens</i> Ehrenberg
Mel var	<i>Melosira varians</i> C. Agardh	Stau goep	<i>Stauroneis goeppertiana</i> Bleisch(Navmutvartrop)
Mer cir	<i>Meridion circulare</i> (Greville) C. Agardh	Stau jav	<i>Stauroneis javanica</i> (Grunow) Cleve
Nav amp	<i>Navicula amphioxys</i> Kutzing	Stau phoe	<i>Stauroneis phoenicenteron</i> (Nitzsch) Ehrenberg
Nav at	<i>Navicula atomus</i> (Kutzing) Grunow	Sur ang	<i>Surirella angusta</i> Kutzing
Nav car	<i>Navicula cari</i> Ehrenberg W. Gregory ex Greville	Sur lin	<i>Surirella linearis</i> W. Smith
Nav cf pla	<i>Navicula cf. placentula</i> (Ehrenberg) Kutzing	Sur ova	<i>Surirella ovalis</i> Brebisson
Nav cf pse	<i>Navicula cf. pseudoscutiformis</i> (Hustedt)	Sur rob	<i>Surirella robusta</i> Ehrenberg
Nav coc	<i>Navicula cocconeiformis</i> W. Gregory ex Greville	Sur ten	<i>Surirella tenera</i> W. Gregory
Nav cons	<i>Navicula constans</i> Hustedt	Syn ac	<i>Synedra acus</i> Kutzing
Nav cryp	<i>Navicula cryptotenella</i> Lange-Bertalot	Syn tab	<i>Synedra tabulata</i> (C. Agardh) Kutzing
Nav cus	<i>Navicula cuspidata</i> (Kutzing) Kutzing	Syn ul	<i>Synedra ulna</i> (Nitzsch) Ehrenberg
Nav daf	<i>Navicula distans</i> W. Smith	Tab fasc	<i>Tabularia fasciculata</i> (C. Agardh) D.M. Williams & Round
Nav elg	<i>Navicula elginensis</i> (W. Gregory) Ralfs	Tab floc	<i>Tabellaria flocculosa</i> (Roth) Kutzing
Nav greg	<i>Navicula gregaria</i> Donkin	Try cal	<i>Tryblionella calida</i> (Grunow) D.G. Mann
Nav has	<i>Navicula hasta</i> Pantocsek		

Table 4.WA and WA tolerance down-weighted (WAT), RMSE, regression of 40 samples for transfer function development

Code	RMSE	R ²	Ave_Bias	Max_Bias	Jack_R ²	Jack_Ave_Bias	Jack_Max_Bias	RMSEP
WA_Inv	0.22	0.50	0.00	0.75	0.04	-0.01	1.16	0.33
WA_Cla	0.31	0.50	0.00	0.31	0.07	0.00	1.11	0.40
WATOL_Inv	0.23	0.46	0.00	0.95	0.03	0.05	1.66	0.40
WATOL_Cla	0.34	0.46	0.00	0.67	0.01	0.11	1.99	0.58

Note: WA_Inv: Weighted averaging model (inverse deshrinking) for Total P. WA_Cla: Weighted averaging model (classical deshrinking) for Total P. WATOL_Inv: Weighted averaging model (tolerance down weighted, inverse deshrinking) for Total P. WATOL_Cla: Weighted averaging model (tolerance down weighted, classical deshrinking) for Total P

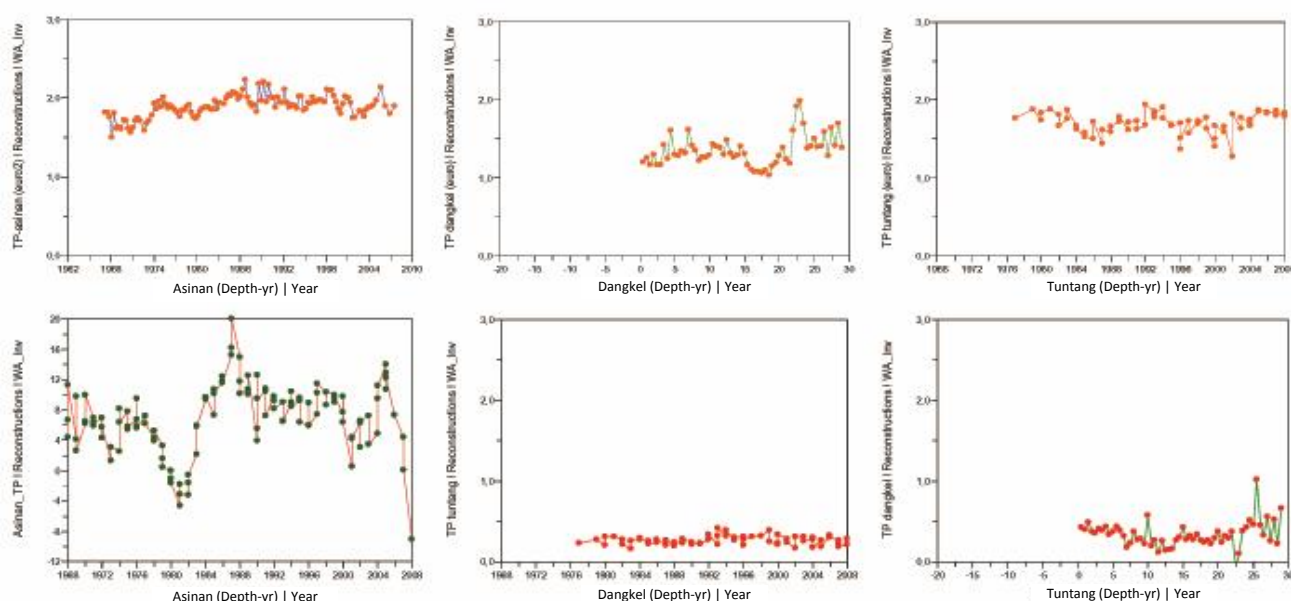


Figure 3. The past total phosphorous reconstruction of Rawapening Lake, Central Java, Indonesia with European diatom data set (above) and Rawapening diatom data set (below)

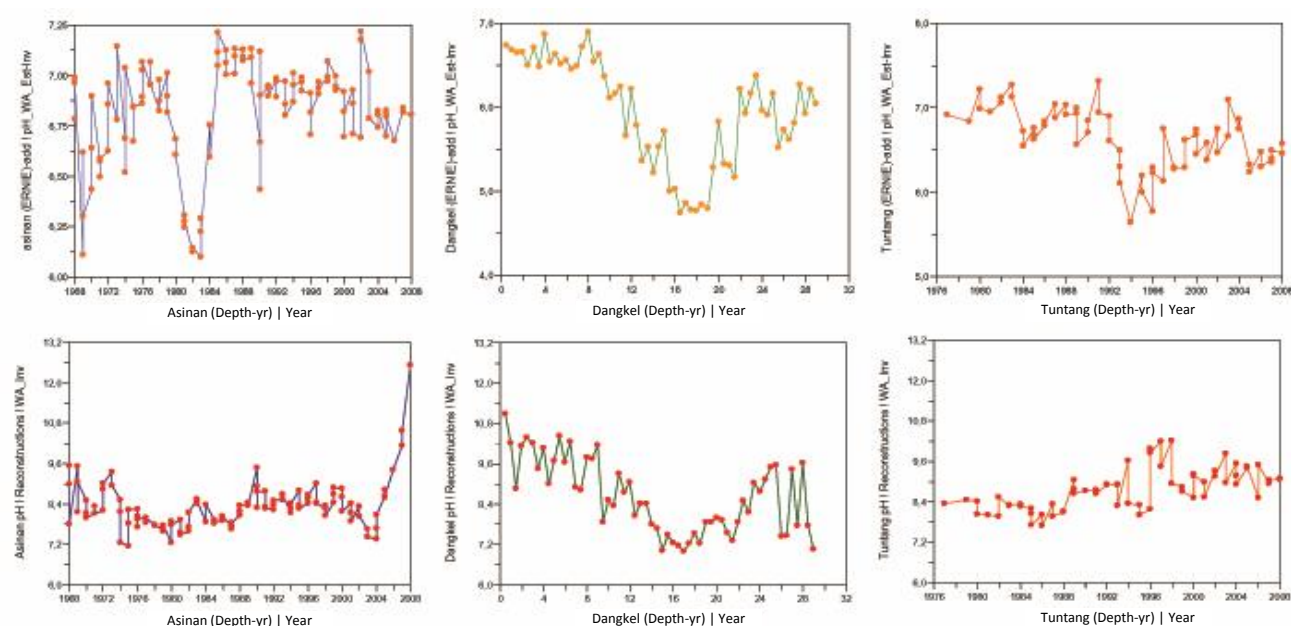


Figure 4. Reconstruction of pH at 3 sites using European data set (above) and Rawapening Lake dataset (below)

The conclusion shows that phosphate, temperature, and calcium were the environmental parameters that influence the diatoms assemblage of Rawapening Lake. Phosphate was contributed 50% on diatoms assemblage. Internal based transfer function of diatoms provides more suitable diatoms data set rather than European diatoms data set. The Rawapening diatom data set of total phosphorous is the initial Indonesian diatom data set for past trophic status, therefore, research has to be continued spatially for other

Indonesian lakes and temporary on specific Indonesian lake to develop Indonesia diatoms data set.

ACKNOWLEDGEMENTS

Thanks for Keely Mills and Rosie Grundell from The Federation University Australia, Ballarat, Australia for helping on the diatoms analysis, Jafron Wasiq Hidayat and

Kariyadi Baskoro from Universitas Diponegoro, Semarang, Indonesia for helping with the fieldworks, and Nina Desianti from Lab Academy of Natural Sciences Drexel University, USA for her help with the data analysis. This article was produced as a part of project supported by Indonesian Higher Education through Doctorate Research Grant for Universitas Gadjah Mada, Contract Number: LPPM-UGM/1177/2009, 19 May 2009, Australian Institute of Nuclear Science and Engineering (AINSE) Grant 09065. The recent data were supported by Competence Research Grant supported by Directorate Research and Community Services, Directorate General of Higher Education, The Ministry of Education and Culture, Year 2014, through DIPA Undip, Contract Number: 023.04.2.189185/2014,03 March 2014.

REFERENCES

- Adams KE, Taranu ZE, Zurawell R, Cumming BF, Eaves IG. 2014. Insights for lake management gained when paleolimnological and water column monitoring studies are combined: A case study from Baptiste Lake. *Lake Reserve Manag* 30 (1): 11-22.
- Avans AEV, Hanjra MA, Jiang Y, Qadir M, Drechsel P. 2012. Water Quality: Assessment of the Current Situation in Asia. *Water Res Dev* 28 (2): 195-216
- Battarbee R, Jones VJ, Flower RJ, Cameron NG, Bennion H, Carvalho L, Juggins S. 2001. Diatoms. In: Smol JP, Birks HJB, Last WM (eds.). *Tracking Environmental Change Using Lake Sediments. Volume 3: Terrestrial, Algal and Silicious Indicators*. Kluwer Academic Publishers, Nederland.
- Birks HJB, Heiri O, Seppa H, Björne AE. 2010. Strengths and Weaknesses of Quantitative Climate Reconstructions Based on Late-Quaternary Biological Proxies. *Open Ecol J* 3: 68-110.
- BPD-L-PPLH Undip. 1999. Environmental Quality Index Development and Biological Indicator. Badan Pengendalian Dampak Lingkungan-Pusat Penelitian Lingkungan Hidup, Universitas Diponegoro, Semarang. [Indonesian].
- BPS [Central Bureau of Statistics]. 2010. 2010 Semarang District in Figure. Badan Pusat Statistik Kabupaten Semarang, Semarang.
- Gell P, Tibby J, Little F, Baldwin D, Hancock G. 2007. The impact of regulation and salinization on floodplain lakes: The lower River Murray, Australia. *Hydrobiologia* 591: 135-146.
- Goltenboth F. 1979. Preliminary final report. The Rawapening Project. Universitas Kristen Satya Wacana, Salatiga.
- Guiry MD, Guiry GM. 2016. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. [9 June 2016]. <http://www.algaebase.org>.
- Juggins S. 2014. User Guide C2. Software for ecological and paleontological data analysis and visualisation. User Guide version 1.7.6. University of New Castle, New Castle, UK.
- Juggins S. 2016. The European Diatom Database User Guide Version 1.0. University of New Castle, New Castle, UK.
- Kireta AR, Reavie ED, Sgro GV, Angradi, TR, Bolgrien DW, Jicha TM, and Hill BH. 2012. Assessing the condition of Missouri, Ohio, and Upper Mississippi rivers (USA) using diatom-based indicators. *Hydrobiologia* 691 (1): 171-188.
- Kramer K, Lange-Bertalot H. 2008. *Subwasserflora Von Mitteleuropa*, Bd. 02/3: Bacillariophyceae: Teil3: Centrales, Fragillariaceae, Eunotiaceae. Spectrum, Berlin.
- Kramer K, Lange-Bertalot H. 2010a. *Subwasserflora Von Mitteleuropa*, Bd. 02/1: Bacillariophyceae: Teil 1: Naviculaceae. Spectrum, Berlin.
- Kramer K, Lange-Bertalot H. 2010b. *Subwasserflora Von Mitteleuropa*, Bd. 02/2: Bacillariophyceae: Teil 2: Bacillariophyceae, Epithemiaceae, Surirellaceae. Spectrum, Berlin.
- Kramer K, Lange-Bertalot H. 2011. *Subwasserflora Von Mitteleuropa*, Bd. 02/4: Bacillariophyceae: Teil 4: Achnanthes S.I., Navicula Str. Spectrum, Berlin.
- Mackay AW, Battarbee RW, Flower RJ. 2003. Assessing the potential for developing internal diatom-based transfer functions for Lake Baikal. *Limnol Oceanogr* 48 (3): 1183-1192.
- Mills K. 2009. Ugandan Crater Lakes Limnology, Palaeolimnology and Palaeoenvironmental History. [Ph.D. Dissertation]. Loughborough University, England.
- MoE [Ministry of Environment]. 2010. 2010-2014 National Lake Priority Program. Ministry of Environment, Jakarta. [Indonesian]
- MoE [Ministry of Environment]. 2011. GERMADAN-Save Rawapening Lake. Ministry of Environment, Jakarta. [Indonesian]
- Reid MA, Ogden RW. 2009. Factor affecting diatom distribution in floodplain lakes of the Southeast Murray basin, Australia and implications for paleolimnological studies. *J Paleol* 41: 453-470.
- Soeprbowati TR, Suedy SWA, Gell P. 2012. Diatom stratigraphy of mangrove ecosystems on the northern coast of Central Java. *J Coastal Dev* 15 (2): 197-208
- Soeprbowati TR. 2010. The standard method of using diatom as earlier warning indicator of water quality changing. Proceeding the International Conference of Management of Innovation and Technology, October 27, 2000. Universitas Diponegoro, Semarang.
- Soeprbowati TR. 2015. Bioindicator of water quality. Proceeding of the National Seminar on Biology, Program Magister Biologi Universitas Diponegoro, Semarang, August 6, 2015.
- ter Braak CJF, Smilauer P. 2009. CANOCO for Window version 4.56. Biometris-Plant Research International, Wageningen, The Netherlands.
- Wibowo H. 2004. Eutrophication of Rawapening Lake and Primary Production of Phytoplankton. [Thesis]. Program of Environmental Science. Graduate Program, Universitas Diponegoro, Semarang.
- Yun SM, Joo HM, Jung SW, Choi CH, Ki JS, Lee JH. 2014. The relationship between epilithic diatom communities and changes in water quality along the lower Han River, South Korea. *J Freshwater Ecol* 29 (3): 363-375.