

# Green synthesis of copper nanoparticles using *Musa acuminata* aqueous extract and their antibacterial activity

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**Abstract.** Muhammad A, Umar A, Birnin-Yauri AU, Sanni HA, Elinge CM, Ige AR, Ambursa MM. 2023. Green synthesis of copper nanoparticles using *Musa acuminata* aqueous extract and their antibacterial activity. *Asian J Trop Biotechnol* 20: 10-16. Green synthesis is a convenient and affordable alternative to the synthesis of copper nanoparticles. The objective of the present study was to synthesize copper nanoparticles via the green method, using *Musa acuminata* (banana) fruit extracts as a reducing agent with copper sulfate serving as a precursor. The antibacterial activities of the synthesized CuNPs were also tested. Fruits extracts of *M. acuminata* were used to synthesize with CuSO<sub>4</sub>(aq) as a precursor. The synthesized nanoparticles were characterized using UV-Vis, FTIR, XRD, SEM, and EDX techniques. The synthesis results showed that the nanoparticles were of Face-Centered Cubic (FCC) structure with high stability. The average diameter of the synthesized copper nanoparticles is 18nm. UV-Vis, scanning electron microscope, X-ray diffraction, energy disperses X-ray, and Fourier transforms infrared spectroscopy established that the formed nanoparticles are copper nanoparticles. The antibacterial activity of the synthesized nanoparticles was also tested using pathogenic bacteria *Escherichia coli* and *Staphylococcus aureus*. The results showed that copper nanoparticles were promising antibacterial agents.

**Keywords:** Antibacterial activity, banana, copper nanoparticles, CuSO<sub>4</sub> solution, *Musa acuminata* extract

## INTRODUCTION

Green synthesis can be defined as the derivation of materials from green or eco-friendly resources by using solvents, excellent reducing agents, and harm less material for stabilization (Bonde 2011; Sable et al. 2012; Jadoun et al. 2021). Furthermore, this synthesis route is simple, economical, reliable, sustainable, and relatively reproducible and provides more stable compounds. Therefore, researchers are interested in developing various nanomaterials through this biosynthesis pathway, including metal/metal oxide nanoparticles, hybrid materials, and biologically inspired materials. As a result, green synthesis is widely regarded as a necessary tool to reduce the negative effects of conventional nanoparticle synthesis methods used in laboratories and industry (Yedurkar et al. 2017).

With numerous reports and studies, this green synthesis process already produces a large number of metal/metal oxide nanoparticles such as silver (Ag), gold (Au), selenium (Se), platinum (Pt), zinc oxide, etc. (Chakraborty et al. 2022). Furthermore, the studies also reported that some metallic nanoparticles have a variety of biological and biochemical activities, but CuNPs have recently received a lot of attention. Copper plays a variety of roles in humans health, including the production of neuropeptides, regulation of cell signaling pathways, antioxidant defense, and co-factoring of many enzymes involved in immune cell function (Georgopoulos et al.

2001; Ghaderian and Ravandi 2012).

Copper Nanoparticles (CuNPs) attract positive global attention because of their low-cost and novel optical, mechanical, catalytic, electrical and thermal conduction properties, which differ from that of bulk metals (Brust and Kiely 2002). Copper nanoparticles have been considered alternatives for noble metals in many applications, such as heat transfer and microelectronics. However, the synthesis of copper nanoparticles poses a challenge due to the high tendency for oxidation. Unlike gold and silver, copper is extremely sensitive to air, and the oxide phases are thermodynamically stable (Jeong et al. 2008).

Plants produce many biologically active chemicals. Because very small amounts of these heavy metals are hazardous even at very low concentrations, plants have shown extraordinary potential in detoxifying and accumulating heavy metals, which may allow them to outrun environmental pollutants (Chakraborty et al. 2022). In addition, the synthesis of nanoparticles using plant extracts is superior to other biological synthesis methods, such as microorganisms. That is because the rate of synthesis of metallic nanoparticles using plant extracts is more sustained (Saif et al. 2016), significantly faster and more monodisperse than other biological methods (Sarkar et al. 2020).

Copper nanoparticles have been synthesized using extracts from various plants found all over the globe (Murthy et al. 2020). From this note, many plant parts, fruits, or whole plants have been used for the green

synthesis of CuNPs due to the presence of a large number of bioactive compounds (Murthy et al. 2020). The Synthesis of CuNPs has been successfully extracted from various parts of *Punica granatum* (Kaur et al. 2016), *Zingiber officinale* stem (Delma and Rajan 2016), *Citrus medica* juice (Shende et al. 2015), *Ziziphus spina-christi* fruit (Khani et al. 2018), *Asparagus adscendens* roots and leaves (Thakur et al. 2018); leaves of *Eclipta prostrata* leaves (Chung et al. 2017); *Ginkgo biloba* leaves (Nasrollahzadeh and Mohammad 2015); *Plantago asiatica* leaves (Nasrollahzadeh et al. 2017); *Thymus vulgaris* (Issaabadi et al. 2017); Black tea leaves (Asghar et al. 2018); *Terminalia catappa* leaves (Muthulakshmi et al. 2017) and *Azadirachta indica* (Ahmed et al. 2015).

This research work synthesized CuNP from copper sulfate pentahydrate and aqueous extract from *Musa acuminata*. Advanced techniques characterized the copper nanoparticles, and an antimicrobial assay was performed.

## MATERIALS AND METHODS

The fruits of *M. acuminata* (banana) were obtained from Sokoto central market for this study. The representative samples for the analysis were selected by random sampling. The samples of the banana fruits were peeled, and 20 g were accurately weighed. The sample was then crushed and finely macerated with mortar and pestle. The sample was then added to 100cm<sup>3</sup> of deionized water and boiled over a water bath at 60°C for 15 minutes. The extract was allowed to cool down and filtered with muslin cloth through Whatman filter paper and used immediately for the synthesis. A 0.01 M solution of CuSO<sub>4</sub>·5H<sub>2</sub>O was prepared by dissolving 2.4955 g of the salt in 10 cm<sup>3</sup> of deionized water.

### Green synthesis of copper nanoparticles

About 10 cm<sup>3</sup> of the *M. acuminata* extract was added to 100 cm<sup>3</sup> of 0.01 M CuSO<sub>4</sub>·5H<sub>2</sub>O aqueous solution and mixed thoroughly. The mixture was heated to 80°C with constant stirring on a magnetic stirrer for six h. The suspension produced was centrifuged at 3,000 rpm for 10 min, and the supernatant liquid was decanted. The residue was repeatedly washed with 10 cm<sup>3</sup> of deionized water. Centrifugation-decantation-washing processes were repeated five times to remove impurities on the surface of the copper nanoparticles. The obtained precipitates were dried in an oven at 50°C for 24 h. The synthesized copper nanoparticles were then kept for characterization and antimicrobial studies.

### Characterization

#### UV-Visible spectroscopy

UV-Visible spectroscopy technique was used in this research to confirm the formation of nanoparticles. For each analysis, 1.0 cm<sup>3</sup> of the aliquot suspension was diluted in 4.0 cm<sup>3</sup> deionized water, and its UV-Visible spectrum was measured at 80°C (Gnanasangeetha and Prathipa 2019).

#### Fourier transform infrared spectroscopy (FTIR)

FTIR analysis was carried out to identify the possible biomolecules responsible for reducing copper sulfate to CuNPs and the capping and stabilizing agents for CuNPs. After synthesizing the copper nanoparticles, the precipitate obtained using each extract was dried in an oven at 50°C for 24 h. The dried synthesized copper nanoparticles were grounded with KBr, cast into a pellet, and used for analysis on the FTIR spectrophotometer in the diffuse reflectance mode operating at a resolution of 4 cm<sup>-1</sup> (Gnanasangeetha and Sarala 2015).

#### X-Ray Diffraction studies (XRD)

X-ray diffraction was used to determine the phase and crystallinity of the nanoparticles. The dried samples of the as-synthesized copper nanoparticles were grounded to fine powder and a thin film of each sample was made by dipping a cleaned glass plate into the powdered nanoparticles of the sample. The XRD analysis was performed passing monochromatic Cu radiation ( $\theta = 1.5406 \text{ \AA}$ ) operating at a voltage of 40kV and a current of 25mA on the sample film at room temperature (25°C). The XRD patterns were then collected at  $2\theta$  angles between 10° and 80°, 0.02 min<sup>-1</sup> and at 1 second time constant. The peaks obtained were compared with the reference database in the Joint Committee on Powder Diffraction Standards (card no: 53-61386) library to ascertain the nature of the nanoparticles (Gnanasangeetha and Sarala 2015).

#### Scanning Electron Microscopy (SEM)

The SEM is a multipurpose instrument that provides qualitative information about the samples, such as topography, morphology, composition, and crystallographic arrangements, with high resolution (Gnanasangeetha and Suresh 2020). First, the surface morphology of the synthesized copper nanoparticles was studied using Scanning Electron Microscopy (SEM). Next, the sample was prepared to study size and crystallographic structure by placing a drop of colloidal solution of the synthesized copper nanoparticles in copper (II) sulfate on a carbon-coated copper slide and subsequently drying it in the air before transferring it to the microscope. The operation was carried out at an accelerated voltage of 130 kV, magnification of X<sup>10</sup>, and resolution of 1 nm (Sarala 2015).

#### Energy Dispersive X-ray (EDX)

The use of EDX analysis investigated the presence of metallic copper. The copper nanoparticles' elemental composition, purity, and geometry were studied. Copper nanoparticles had a sorption peak at 1Kev (Ebrahimi et al. 2017), the point at which the index for metallic nanoparticles of copper was expected. The EDX observation was carried out by instrument coupled with SEM (Gnanasangeetha and Sarala 2015).

#### Test of antibacterial activity

The antibacterial activity of the synthesized copper nanoparticles was investigated according to the method outlined by Ebrahimi et al. (2017). The assays were

conducted on agar well diffusion method with Gram-negative bacteria *Escherichia coli* and Gram-positive bacteria *Staphylococcus aureus*. The pathogenic bacteria were cultured using nutrient agar in petri dishes with an inner diameter of 9 cm to provide a thin agar plate of thickness 3.4-3.5 mm after solidification. The culture was regulated to 0.5 McFarland standards to get  $1.5 \times 10^6$  CFU/mL (Ebrahimi et al. 2017).

The sample was prepared for an antibacterial test and labeled as A for the banana (*M. acuminata*) mediated Cu nanoparticles. The extracts were prepared at 1 mg/mL and dissolved in Dimethyl Sulfoxide (DMSO) in each case (Murthy et al. 2020). A Hollow of 6mm diameter was cut from each assay using a sterile cork-borer, and 50  $\mu$ L of each of the three extracts was impregnated using a sterilized wire loop on the surface of Mueller-Hinton Agar (MHA) plates. The pathogens were incubated at 5-8°C for 2 hours to ensure good diffusion and then further incubated at 37°C for 24 hours (Ebrahimi et al. 2017). Ampicillin disc was used as a control. Next, using a micropipette, 10 $\mu$ L of antibiotic control (Ampicillin) was measured and used for both the *E. coli* and the *S. aureus* bacteria (Ebrahimi et al. 2017). The diameters of the inhibition zone in each case were measured millimeters using a ruler, and the results were recorded.

## RESULTS AND DISCUSSIONS

### UV-Visible absorption spectroscopic study

UV-Visible spectra of aqueous  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  mediated with the *M. acuminata* fruits extract are shown in Figure 1. The reduction of  $\text{Cu}^{2+}$  ions to Cu nanoparticles by the sample extract was indicated by the change of color from pale yellow to reddish brown.

The progress in reducing  $\text{Cu}^{2+}$  ions to Cu nanoparticles was indicated by the enhanced intensity of surface Plasmon absorption peak observed within 560nm to 600nm. The maximum absorption observed was 574nm. The appearance of the peak assigned to the Surface Plasmon Resonance band is within the range of 550nm-600nm for copper nanoparticles, as reported by Khodaie and Ghasemi (2018).

The metal nanoparticles show strong absorption of visible radiation due to their induced polarization in their conduction electrons concerning the immobile nucleus (Michael 2012). Furthermore, when a particular wave length is matched to the size of a nanoparticle, dipole oscillation is generated in the compensated form of the induced polarization, and the electrons in the nanoparticle resonate, introducing a strong absorption (Moskovits and Vlckova 2005).

### Fourier Transform Infrared spectroscopic study (FTIR)

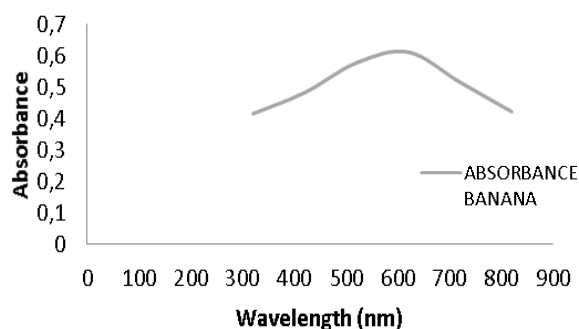
FTIR spectroscopic measurement was conducted to discover the possible biomolecules in the *M. acuminata* (banana) fruit extract and the synthesized Cu nanoparticles. Figure 2 showed active functional groups in the fruit extracts and the functional groups in the synthesized

CuNPs. The figures showed a comparative Fourier Transform Infrared (FTIR) spectroscopic analysis of sample extracts and the synthesized CuNPs.

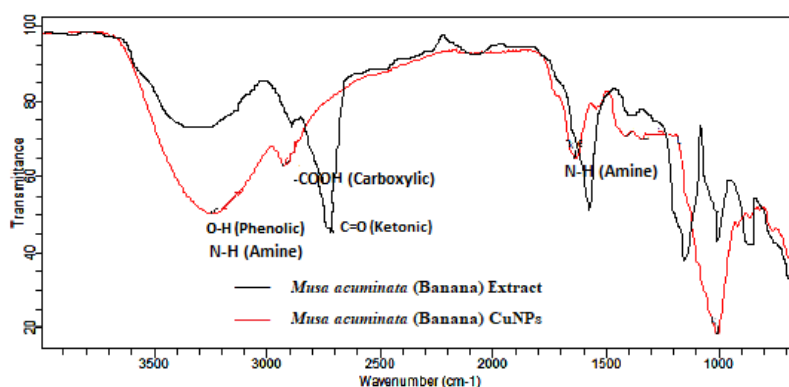
Peaks observed around  $3,200 \text{ cm}^{-1}$  and  $2,900 \text{ cm}^{-1}$  are characteristics of a carboxylic COOH (or N-H stretching mode) and alkynic  $\equiv\text{C-H}$  stretching. Peaks at  $1,700 \text{ cm}^{-1}$ ,  $1,600 \text{ cm}^{-1}$  and  $1,618 \text{ cm}^{-1}$  are corresponding to amide, arising due to carbonyl stretching in proteins, and the band at  $1,604 \text{ cm}^{-1}$  is a characteristic of N-H bending. The peaks at  $1,474$  and  $1,039-1,381 \text{ cm}^{-1}$  correspond to methylene scissoring vibrations from the proteins in the solution and C-N stretching vibrations of amine (Michael 2012). The FTIR spectra showed that the flavonoids, alkaloids, protein molecules, and other metabolites in the fruit extract are responsible for reducing copper ions and stabilizing the Cu nanoparticles. The FTIR data were in agreement with the previous reports (e.g., Dadgostar 2008; Hailemariam 2011; Valodkar et al. 2011; Kalainila et al. 2014; Gnanasangeetha and Prathipa 2019). It is also clear from the relative FTIR data that the peaks of the functional groups of the spectra of the fruit extract and those of their corresponding synthesized copper nanoparticles are almost similar. That signifies the impact of the functional groups in the fruit extracts in reducing  $\text{Cu}^{2+}$  to CuO and further stabilization of CuNPs, as stated by Chandraker et al. (2020).

### X-Ray Diffraction studies (XRD)

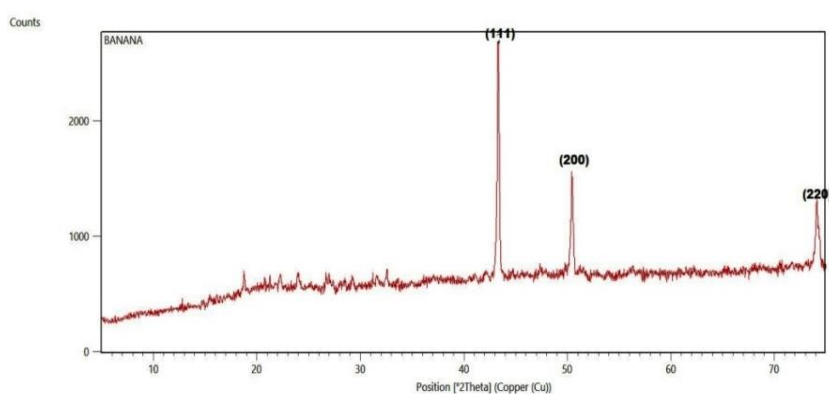
The XRD pattern of the synthesized Cu nanoparticles obtained by the green reduction of copper ions using banana (*M. acuminata*) fruit extracts is presented in Figure 3. The values of experimental diffraction peaks observed in the patterns at  $2\theta$  for CuNPs synthesized from banana fruit extract are 43.50, 50.40, and 74.10. These values of diffraction data conformed with Inorganic Structure Data Base (ICSD); file no. 04-0836 (Rajesh et al. 2018) as well as with International Centre for Diffraction Data (ICDD) standard for value for CuNPs; file number 04-0836 (Theivasanthi and Alagar 2011). The three distinct diffraction peaks for the copper nanoparticles synthesized from the banana correspond to (111), (200), and (220) lattice planes of the face-centered cubic structure (FCC) of copper nanoparticles.



**Figure 1.** UV-Visible spectra of the synthesis of copper nanoparticles for the three fruits extracts



**Figure 2.** FTIR spectra of *Musa acuminata* (banana) extract and CuNPs synthesized using *M. acuminata* extract



**Figure 3.** XRD Pattern of CuNPs synthesized using *Musa acuminata* (banana)

### Scanning Electron Microscopy (SEM)

Figure 4 presents an image obtained from Scanning Electron Microscopy (SEM) of the copper nanoparticles synthesized from banana. Figure 4 presents SEM images of copper nanoparticles synthesized from bananas, which showed dispersed copper nanoparticles with irregular edges. The diameters of the copper nanoparticles were 18 nm for the banana, and the shapes of the CuNPs synthesized from banana synthesized nanoparticles appeared dispersed. The results were in agreement with Jaehoon et al. (2006) and Arya et al. (2018).

### Energy Dispersive X-ray (EDX)

The use of EDX analysis confirmed the presence of metallic copper. The observed chart for Energy Dispersive X-ray (EDX) for as-synthesized copper nanoparticles is illustrated in Figure 5.

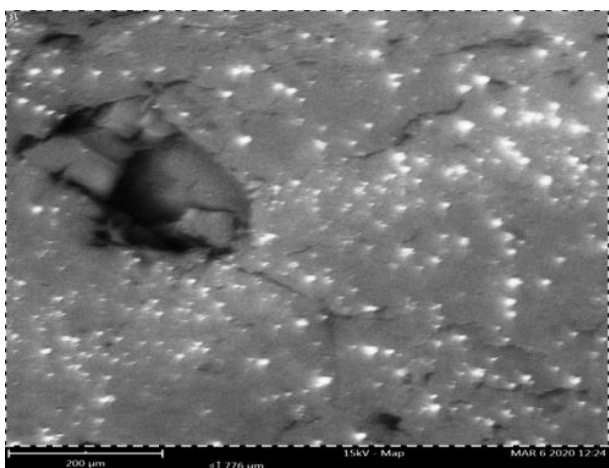
The EDX spectra of the copper nanoparticles synthesized are presented in Figure 5. The EDX spectra confirmed the presence of metallic copper and other elements, which corroborated the findings of Gnanasangeetha and Sarala (2015). The presence of elements such as C, S, and Cl may be attributed to contaminations originating from the biomolecules bound to the surface of the copper nanoparticles or physical absorption during sample preparation. In addition, oxygen

and the Cu signal signify that phytoconstituents capped the CuNPs through oxygen atoms. In contrast, the presence of a trace amount of carbon demonstrated the involvement of plant phytochemical groups in the reduction and capping of the synthesized CuNPs.

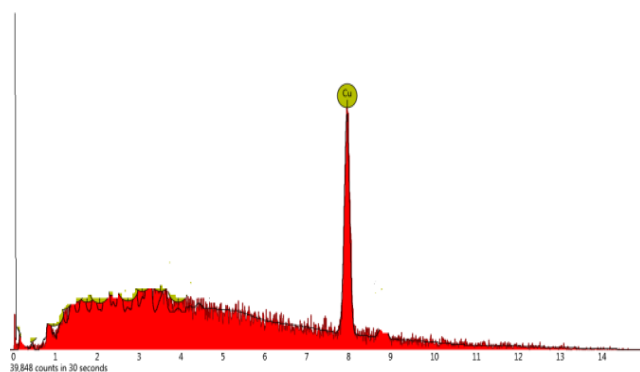
### Antibacterial activities of copper nanoparticles

The antibacterial activities of banana (*M. acuminata*) fruits extract mediated copper nanoparticles against two pathogenic bacteria, Gram-negative *E. coli* and Gram-positive *S. aureus* using agar well diffusion method shown in Table 1. Figure 6 illustrated the antibacterial activity of *M. acuminata* synthesized CuNps and Ampicillin, respectively, on *S. aureus* and *E. coli* bacteria.

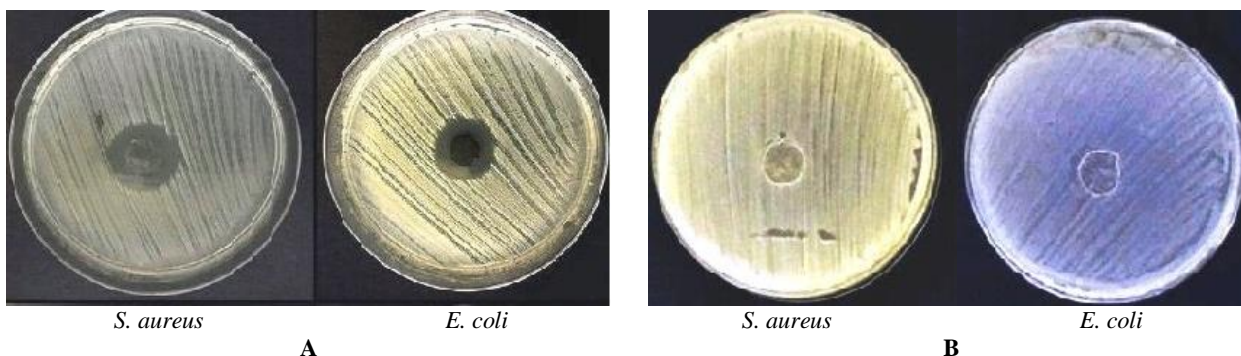
The mean values of three replicates of the zone of inhibition (mm) around each assay of the synthesized copper nanoparticles are presented in Table 1. The results of antibacterial activities of banana (*M. acuminata*) fruits extract mediated copper nanoparticles against two pathogenic bacteria, Gram-negative *E. coli* and Gram-positive *S. aureus* using agar well diffusion method and are presented in Table 1. The diameter of the assays' inhibition zones was measured in millimeters (mm) using a ruler. The results were recorded and compared to those obtained from the standard antibiotic drug, Ampicillin.



**Figure 4.** SEM image of CuNPs synthesized using *Musa acuminata* (banana)



**Figure 5.** EDX figure of CuNPs synthesized using *Musa acuminata* (banana)



**Figure 6.** A. The antibacterial activity of *Musa acuminata* synthesized CuNPs (A) and Ampicillin (B), respectively, on *S. aureus* and *E. coli* bacteria

**Table 1.** Mean values of three replicates of the zone of inhibition (mm) around the sample of the synthesized copper nanoparticles

Culture (bacteria)	Diameter of zone of inhibition (mm)	
	<i>Musa acuminata</i> (banana)	Ampicillin (Standard)
<i>Escherichia coli</i>	16±2.2	17±0.50
<i>Staphylococcus aureus</i>	19±2.5	19±1.20

Note: The zone of inhibition was measured in mm. Values are expressed as Mean±SD, where n=3

From the obtained results, copper nanoparticles were found to have higher antibacterial activity against Gram-positive bacteria *S. aureus* than Gram-negative *E. coli*. The antibacterial activity was also more pronounceable against *S. aureus* compared to Ampicillin. The inhibition zones observed for *M. acuminata* synthesized CuNPs are 19±2.5 for *S. aureus* and 16±2.2 for *E. coli* and Ampicillin 19 mm and 17 mm are the values recorded for *S. aureus* and *E. coli*, respectively.

The higher activity against Gram-positive may be attributed to a greater abundance of amines and carboxyl groups on the cell surface of the Gram-Positive Bacteria and a greater affinity of copper towards these groups. According to Rupareli et al. (2008), copper ions released may interact with DNA molecules and insert between the nucleic acid strands. Moreover, copper ions inside bacterial cells also disrupt biochemical processes. The study showed that green synthesized copper nanoparticles from bananas have great potential in biomedical applications. That following with the previous observations of other researchers (Hailemariam 2011; Theivasanthi and Alagar 2011; Joseph et al. 2016).

In conclusion, this research confirmed the feasibility of green synthesis of copper nanoparticles using extracts of *M. acuminata* (banana) fruits. The results indicated a reduction of copper ions and stabilization of copper nanoparticles occurred due to the presence of proteins and other metabolites in the fruit extracts. The size of synthesized CuNPs calculated using powder XRD pattern is 18nm. FT-IR results confirmed that extracts of *M. acuminata* (banana) fruit are a suitable green pattern to

prepare copper nanoparticles. It was also observed that *M. acuminata* (banana) copper nanoparticles were found to have a very good and fabulous antibacterial activity.

## REFERENCES

- Ahmed S, Saifullah M, Ahmad B, Swami L, Ikram, S. 2015. Green synthesis of silver nanoparticles using *Azadirachta indica* aqueous leaf extract. *J Radiat Res Appl Sci* 9 (1): 1-7. DOI: 10.1016/j.jrras.2015.06.006.
- Arya, A, Gupta, K, Chundawat, TS, Vaya D. 2018. Biogenic synthesis of copper and silver nanoparticles using green alga *Botryococcus braunii* and its antimicrobial activity. *Bioinorg Chem Appl* 9: 7879403.
- Asghar MA, Zahir E, Shahid SM. 2018. Iron, copper and silver nanoparticles: green synthesis using green and black tea leaves extracts and evaluation of antibacterial, antifungal and aflatoxin B<sub>1</sub> adsorption activity. *LW* 90: 98-107.
- Bonde S. 2011. A biogenic approach for green synthesis of silver nanoparticles using extract of *Foeniculum vulgare* and its activity against *Staphylococcus aureus* and *Escherichia coli*. *Nusantara Biosci* 3: 59-63. DOI: 10.13057/nusbiosci/n030201.
- Brust M, Kiely CJ. 2002. Some recent advances in nanostructure preparation from gold and silver particles: A short topical review. *Colloids Surf A: Physicochem Eng Asp* 202 (2-3): 175-186. DOI: 10.1016/S0927-7757(01)01087-1.
- Chakraborty N, Banerjee J, Chakraborty P, Banerjee A, Chanda C, Ray K. 2022. Green synthesis of copper/copper oxide nanoparticles and their applications: A review. *15: 187-215*. DOI: 10.1080/17518253.2022.2025916.
- Chandraker SK, La M, Ghosh MK, Tiwari V, Ghorai TK, Shukla R. 2020. Green synthesis of copper nanoparticles using leaf extract of *Ageratum houstonianum* Mill. and study of their photocatalytic and antibacterial activities. *Nano Express* 1: 010033. DOI: 10.1088/2632-959X/ab8e99.
- Chung IM, Rahuman A, Marimuthu S. 2017. Green synthesis of copper nanoparticles using *Eclipta prostrata* leaves extract and their antioxidant and cytotoxic activities. *Exp Ther Med* 14: 18-24. DOI: 10.3892/etm.2017.4466.
- Dadgostar N. 2008. Investigations on Colloidal Synthesis of Copper Nanoparticles in a Two-phase Liquid-liquid System. [Thesis]. University of Waterloo, Waterloo, Ontario. [Canada].
- Delma MT, Rajan MJ. 2016. Green synthesis of copper and lead nanoparticles using *Zingiber officinale* stem extract. *Intl J Sci Res Pub* 6: 134-137.
- Ebrahimi K, Shiravand H, Mahmoudvand H. 2017. Biosynthesis of copper nanoparticles using aqueous extract of *Capparis spinosa* fruit and investigation of its antibacterial activity. *MARMARA Pharm J* 21 (4): 866-871. DOI: 10.12991/mpj.2017.31.
- Georgopoulos PG, Roy A, Yonone-Lioy MJ, Opiokun RE, Lioy PJ. 2001. Environmental copper: Its dynamics and human exposure issues. *J Toxicol Environ Health B* 4: 341-394. DOI: 10.1080/109374001753146207.
- Ghaderian SM, Ravandi AAG. 2012. Accumulation of Copper and other heavy metals by plants growing on Sarcheshmeh Copper Mining Area, Iran. *J Geochem Expl* 2012: 25-32. DOI: 10.1016/J.GEXPLO.2012.06.022.
- Gnanasangeetha D, Prathipa V. 2019. Zinc oxide nanoparticles for water remediation in agriculture. *Intl J Mech Eng Technol* 10 (1): 1547-1554.
- Gnanasangeetha D, Sarala D. 2015. Green synthesis of benovolent ZnO nanods using *Emblica officinalis*. *Asian J Chem* 27 (8): 3054-3056. DOI: 10.14233/ajchem.2015.18870.
- Gnanasangeetha, D, Suresh M. 2020. A review on green synthesis of metal and metal oxide nanoparticles. *Nat Environ Pollut Technol Intl Quart Sci J* 19 (5): 1789-1800 DOI: 10.46488/NEPT.2020.v19i05.002.
- Hailemariam G. 2011. Kinetic Study of Silver Ions Bioreduction for the Synthesis of Silver Nanoparticles and their Antibacterial Activity. [Thesis]. Haramaya University, Haramaya. [Ethiopia]
- Issaabadi Z, Nasrollahzadeh M, Sajadi MS. 2017. Green synthesis of the copper nanoparticles supported on bentonite and investigation of its catalytic activity. *J Clean Prod* 1: 3584-3591. DOI: 10.1016/j.jclepro.2016.10.109.
- Jadoun S, Arif R, Jangid NK, Meena RK. 2021. Green synthesis of nanoparticles using plant extracts: A review. *Environ Chem Lett* 19: 355-374. DOI: 10.1007/s10311-020-01074-x.
- Jaehoon L, Kim DK, Kang W. 2006. Preparation of CII Nanoparticles from CII Powder Dispersed in 2-Propanol by Laser Ablation. *Bull Kor Chem Soc* 27: 1869. DOI: 10.5012/bkcs.2006.27.11.1869.
- Jeong J, Woo S, Kim D, Lim S, Kim JS, Shin H, Xia Y. 2008. Moon controlling the thickness of the surface oxide layer on Cu nanoparticles for the fabrication of conductive structures by ink-jet printing. *Adv Func Mater* 18 (5): 679-686. DOI: 10.1002/adfm.200700902.
- Joseph AT, Prakash P, Narvi SS. 2016. Phytofabrication and characterization of copper nanoparticles using *Allium sativum* and its antibacterial activity. *Intl J Sci Eng Technol* 4: 463-472.
- Kalainila P, Subha V, Ravindran RSE, Sahadevan R. 2014. Synthesis and characterization of silver nanoparticle from *Erythrina indic*. *Asian J Pharm Clin Res* 7 (2): 39-43.
- Kaur P, Thakur R, Chaudhury A. 2016. Biogenesis of copper nanoparticles using peel extract of *Punica granatum* and their antimicrobial activity against opportunistic pathogens. *Green Chem Lett Rev* 9: 33-38. DOI: 10.1080/17518253.2016.1141238.
- Khani R, Roostaei B, Bagherzade G, Moudi M. 2018. Green synthesis of copper nanoparticles by fruit extract of *Ziziphus spina-christi* (L.) Willd.: Application for adsorption of triphenylmethane dye and antibacterial assay. *J Mol Liq* 255: 541-549. DOI: 10.1016/j.molliq.2018.02.010.
- Khodaie M, Ghasemi N. 2018. Green synthesis and characterization of copper nanoparticles using *Eryngium campestre* leaf extract. *Bulgarian Chem Commun* 50: 244-250.
- Michael GEH. 2012. Biosynthesis of Copper Nanoparticles Using Some Plant Leaf Extracts, their Characterization and Antibacterial Activity. [Thesis]. Haramaya University. Haramaya. [Ethiopia]
- Moskovits M, Vlckova B. 2005. Adsorbate-induced silver nanoparticle aggregation kinetics. *J Phys Chem B* 109 (31): 14755-14758. DOI: 10.1021/jp051177o.
- Murthy HC, Desalegn A, Kassa T, Abebe B, Assefa T. 2020. Synthesis of green copper nanoparticles using medicinal plant *Hagenia abyssinica* (Brace) JF. Gmel. leaf extract: Antimicrobial properties. *J Nanomater* 2020: 3924081.
- Muthulakshmi L, Rajini N, Nellaiah H, Kathiresan T, Jawaid M, Rajulu AV. 2017. Preparation and properties of cellulose nanocomposite films with in situ generated copper nanoparticles using *Terminalia catappa* leaf extract. *Intl J Biol Macromol* 95: 1064-1071. DOI: 10.1016/j.jbiomac.2016.09.114.
- Nasrollahzadeh M, Mohammad SS. 2015. Green synthesis of copper nanoparticles using *Ginkgo biloba* L. leaf extract and their catalytic activity for the Huisgen [3 + 2] cycloaddition of azides and alkynes at room temperature. *J Colloid Interf Sci* 457: 141-147. DOI: 10.1016/j.jcis.2015.07.004.
- Nasrollahzadeh M, Momeni SS, Sajadi SM. 2017. Green synthesis of copper nanoparticles using *Plantago asiatica* leaf extract and their application for the cyanation of aldehydes using K<sub>4</sub>Fe(CN)<sub>6</sub>. *J Colloid Interf Sci* 506: 471-477. DOI: 10.1016/j.jcis.2017.07.072.
- Rajesh KM, Ajitha B, Reddy AK, Suneetha Y, Reddy PR. 2018. Assisted green synthesis of copper nanoparticles using *Syzygium aromaticum* bud extract: Physical, optical and antimicrobial properties. *Optik* 154: 593-600. DOI: 10.1016/j.ijleo.2017.10.074.
- Rupareli JP, Chatterjee AK, Duttagupta SP, Mukherji S. 2008. Strain specificity in antimicrobial activity of silver and copper nanoparticles. *Acta Biomaterialia* 4: 707-771. DOI: 10.1016/j.actbio.2007.11.006.
- Sable N, Gaikwad S, Bonde S, Gade A, Rai M. 2012. Phytofabrication of silver nanoparticles by using aquatic plant *Hydrilla verticillata*. *Nusantara Biosci* 4: 45-49. DOI: 10.13057/nusbiosci/n040201.
- Saif S, Tahir A, Chen Y. 2016. Green synthesis of iron nanoparticles and their environmental applications and implications. *Nanomaterials* 6: 209. DOI: 10.3390/nano6110209.
- Sarala P. 2015. Surficial geochemical exploration methods. In: Mayer WD, Lahtinen R, O'Brien H (Eds.). *Mineral Deposits of Finland*. Elsevier, Amsterdam. DOI: 10.1016/B978-0-12-410438-9.00027-3.
- Sarkar J, Chakraborty N, Chatterjee A, Bhattacharjee A, Dasgupta D, Acharya K. 2020. Green synthesized copper oxide nanoparticles ameliorate defence and antioxidant enzymes in *Lens Culinaris*. *Nanomaterials* 2020: 10. DOI: 10.3390/nano10020312.

- Shende S, Ingle AP, Gade A, Rai M. 2015. Green synthesis of copper nanoparticles by *Citrus medica* Linn. (Idilimbu) juice and its antimicrobial activity. *World J Microbiol Biotechnol* 31: 865-873. DOI: 10.1007/s11274-015-1840-3.
- Thakur S, Sharma S, Rai R. 2018. Green synthesis of copper nanoparticles using *Asparagus adscendens* Roxb. root and leaf extract and their antimicrobial activities. *Intl J Curr Microbiol Appl Sci* 7 (4): 683-694. DOI: 10.20546/ijcmas.2018.704.077.
- Theivasanthi T, Alagar M. 2011. Nano sized copper particles by electrolytic synthesis and characterizations. *Intl J Phys Sci* 6 (15): 3726-3735.
- Valodkar M, Jadeja RN, Thounaojam MC, Devkar RV, Thakorea S. 2011. Biocompatible synthesis of peptide capped copper nanoparticles and their biological effect on tumor cells. *Mater Chem Phys* 128: 83-89. DOI: 10.1016/j.matchemphys.2011.02.039.
- Yedurkar S, Maurya C, Mahanwar PA. 2017. Biological approach for the synthesis of copper oxide nanoparticles by *Ixora coccineas* leaf extract. *J Mater Environ Sci* 2017: 1173-1177.